Hindawi International Journal of Dentistry Volume 2021, Article ID 5589664, 7 pages https://doi.org/10.1155/2021/5589664



Research Article

Occurrence of Candida albicans in Periodontitis

Brahim Jabri , ¹ Maryem Iken , ² Mohamed Achmit , ³ Sana Rida , ⁴ and Oum Keltoum Ennibi , ⁵

Correspondence should be addressed to Oum Keltoum Ennibi; o.ennibi@um5s.net.ma

Received 6 January 2021; Revised 22 April 2021; Accepted 22 May 2021; Published 29 May 2021

Academic Editor: Lucio Goncalves

Copyright © 2021 Brahim Jabri et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Periodontal diseases are the result of an imbalance between the microbiota and immune defense. The role of yeast in the pathogenesis of these diseases has been studied. This study aims to assess the occurrence of Candida albicans in periodontitis. Materials and Methods. Fifty subjects were recruited for the study (15 healthy individuals and 35 periodontitis subjects). The periodontal examination and plaque sampling were carried out for all patients. Candida albicans identification was based on culture, direct examination, and polymerase chain reaction. The statistical analysis was performed by SPSS 20 (SPSS Inc., Chicago, IL, USA). Results. Twenty percent of the diseased group harbored Candida albicans which was slightly higher than in the healthy group (7%), suggesting that, under normal conditions, yeast does not grow easily in subgingival sites. However, no significant difference between the healthy and periodontitis groups (p = 0.23) was found. Our results also indicated that the presence of Candida albicans was neither gender nor age related in the studied groups. Conclusion. The results of this study suggest that Candida albicans occurs in periodontitis. More studies are needed to clarify the potential role of this yeast in different stages and forms of the disease.

1. Introduction

Among Candida species, Candida albicans is the most widespread yeast associated with healthy and pathologic oral conditions [1, 2]. Indeed, this opportunist microorganism belongs to commensal microflora in the healthy human digestive tract, but can become pathogenic under the influence of general or local favorable factors.

Periodontitis is a worldwide oral disease with a very complex etiopathogenesis, including dysbiosis and host immune responses that comaintain conditions for the occurrence of periodontal disease in susceptible individuals [3, 4]. Clinically, periodontitis involves attachment loss around teeth, forming periodontal pockets, and bone destruction [5, 6]. Several studies have raised the heterogeneity

of the microflora associated with periodontal disease. Indeed, various microorganism species, including Gramnegative anaerobic bacteria organized in a complex biofilm, have been associated with the initiation and progression of periodontitis [7]. Porphyromonas gingivalis, Tannerella forsythia, and Treponema denticola, known as the "red complex" together with Aggregatibacter mycetemcomitans have been identified as the most periopathogenic bacteria associated with different forms of periodontitis worldwide [7, 8]. Lately, some studies discussed also the possible role of viruses [9, 10] and yeasts in periodontitis pathogenesis [11, 12]. Thus, as the disease breaks through, the periodontal pockets become the receptacle of a large number of microorganisms including C. albicans [6, 13, 14]. Nevertheless, the role of this yeast in

¹Research Laboratory in Oral Biology and Biotechnology, Faculty of Dental Medicine of Rabat, Mohammed V University in Rabat, Rabat, Morocco

²Clinical Biology Department, Faculty of Medicine and Pharmacy, Mohammed V University in Rabat, Rabat, Morocco

³Laboratory of Virology, Microbiology and Quality/ Eco-toxicology and Biodiversity,

Faculty of Sciences and Techniques Mohammedia, University Hassan II Casablanca, Casablanca, Morocco

⁴Department of Conservative Dentistry, Faculty of Dental Medicine of Rabat, Mohammed V University in Rabat, Rabat, Morocco ⁵Department of Periodontology, Faculty of Dental Medicine of Rabat, Mohammed V University in Rabat, Rabat, Morocco

the pathogenesis of periodontal disease remains unclear. The aim of the present study is to assess the presence of *Candida albicans* in periodontitis.

2. Materials and Methods

2.1. Study Population. Fifty subjects were recruited from the clinical department of Periodontology, "Center of Consultation and Dental Treatment," Ibn Sina University Hospital of Rabat, Morocco (CCTD-CHIS, Rabat).

The approval for the study was obtained from the Research Ethics Committee of Mohammed V University, Rabat, Morocco (Ethical agreement number 102/19). All subjects were informed and signed the informed consent.

The inclusion criteria included patients in good general health, aged 18 years and above, with at least 20 teeth present, and who have not received any periodontal treatment or antibiotic medication during at least the previous six months prior to the study. The exclusion criteria included patients under orthodontic treatment, those who smoke or chew any other kind of tobacco, diabetic patients, those having symptomatic oral candidosis and/or had been under antifungal treatment, and pregnant and lactating women.

2.2. Methods

2.2.1. Periodontal Examination. Prior to the periodontal examination, all patients signed the informed consent forms. The assessed clinical variables were modified gingival index (GI), plaque index (PI) [15], probing pocket depth (PPD), and clinical attachment loss (CAL). Probing pocket depth and attachment loss were measured using a standard periodontal probe (Hu-friedy, Chicago, IL), at six sites per tooth, i.e., distobuccal, buccal, mesiobuccal, distolingual, lingual, and mesiolingual, in all teeth excluding third molars. A set of full-mouth standardized intraoral radiographs was obtained from each patient.

Based upon clinical parameters, radiograph information, and patient age, we came to the clinical diagnosis based on the classification of the American Academy of Periodontology (AAP) [16].

Thus, in the healthy periodontium, there was no attachment loss, no periodontal pocket, no bone loss, and periodontal probing was ≤ 3 mm, whereas periodontitis was defined as having interdental clinical attachment lost detectable on at least 2 nonadjacent teeth or the presence of buccal or oral CAL \geq 3 mm with pocketing \geq 3 mm detectable at least in 2 teeth, but not associated with non-periodontitisrelated causes (i.e., deep dental caries, CAL on a distal site of a second molar and associated with malposition or resulting from a third molar extraction, endodontic lesion draining through the marginal periodontium, and a vertical root fracture). Furthermore, periodontitis patients were classified by stage and grade based on the classification. Stages were defined based on severity (primarily periodontal breakdown and periodontitis-associated tooth loss) and complexity of management (pocket depth, infrabony defects, tooth mobility, furcation status, and masticatory disturbance). Grades of periodontitis were estimated based on direct or indirect

evidence of progression rate in three categories: slow (grade A), moderate (grade B), and rapid progression (grade C).

2.2.2. Plaque Sampling. A pool of subgingival biofilm samples were collected from four sites (one site per quadrant) in each patient. We selected the deepest site in each quadrant with a pocket depth equal or greater than 5 mm (PPD \geq 5 mm), clinical attachment loss \geq 2 mm, and which were bleeding on probing.

Plaque sampling was obtained by introducing a sterilized absorbent paper point (30 mm diameter) into the gingival sulcus for 20 seconds (SURE DENT CORPORATION; CEO 197). The samples were placed in 2 mL of sterile phosphate buffered saline (PBS) as a transport medium and immediately transferred to the laboratory for analysis.

2.2.3. Yeast Culture. Samples were first processed and cultured. They were dispersed and plated onto a sabouraud chloramphenicol medium for the isolation of yeasts. Plates were incubated at 37°C in non-CO₂ atmosphere for 24 to 48 hours [17, 18]. The plates were checked daily for yeast growth.

Identification was based on colony and cellular morphology. *Candida albicans* colonies on the Sabouraud chloramphenicol medium are creamy whitish and smooth [19] (Figure 1).

Under direct examination: *Candida* yeasts appear as oval, ovoid, or elongated and possibly budding elements [20] (Figure 2).

Pure cultures of yeast colonies were identified and confirmed later as *Candida albicans* by PCR.

2.2.4. Identification of Candida albicans by PCR. Genomic DNA was extracted as follows: 4 to 6 yeast colonies were suspended in 1 ml of sterile 0.9% NaCl. The extraction was performed using a commercially available BIOLINE DNA extraction kit (ISOLATE II GENOMIC DNA KIT, USA) according to the manufacturer's instructions. The microtubes were centrifuged at 5000 g for 10 min. Then, the supernatant was removed and washed once with 2 mM EDTA (pH 8) and centrifuged for 10 min at 5000 g. The supernatant was discarded while the resulting pellet was stored. The extracted DNA was stored at -20°C for later use.

Molecular identification of the identified and purified yeast strains was accomplished by using the following specific primers for *C. albicans* detection, generating an amplicon of 500 base pairs (forward: 5'-TGCTTCAGTGTCAGTTATACCT-3', Reverse: 5'-ACTGCTCAAACCATCTCTGG-3') [20].

PCR amplification was performed in a $25 \,\mu\text{L}$ reaction mixture containing the abovementioned primers ($10 \,\mu\text{M}$ each), $100 \,\text{ng}$ of extracted DNA, $100 \,\mu\text{M}$ of each of the four deoxyribonucleoside triphosphates (dATP, dCTP, dGTP, and dTTP), $0.5 \,\text{U}$ of taq DNA polymerase, and $5 \,\text{mM}$ of buffer.

The PCR amplification parameters were as follows: an initial denaturation of 1 min at 95°C, followed by 35 cycles of denaturation of 15 seconds at 95°C, a hybridization of 20

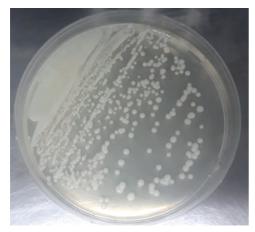


FIGURE 1: Macroscopic appearance of the *Candida* species on the sabouraud chloramphenicol culture medium.

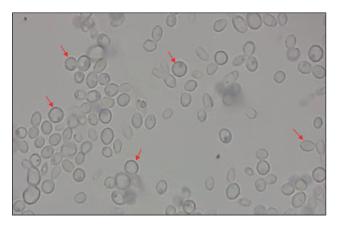


FIGURE 2: Appearance of yeast in a fresh culture (×40).

seconds at 50°C, and an elongation of 15 seconds at 72°C, with a final elongation of 3 min at 72°C. The PCR products were analyzed by 1% agarose gel electrophoresis in the presence of a Dna Ladder 200 bp (Bioline) and visualized by using a UV transilluminator system G-Box (SyngeneTM).

2.2.5. Statistical Analysis. The statistical analysis was performed with a statistical program SPSS 20 (SPSS Inc., Chicago, IL, USA). For each group, continuous variables with normal distribution were presented as mean \pm standard deviation (age, depths of periodontal pockets, and attachment loss), and categorical variables were given as percentage. Chi-square Pearson's test was performed to compare the variables between two independent groups, and Student's t test was performed to compare the means of the independent samples. The statistical significance threshold used was p < 0.05.

3. Results

Fifty subjects accepted to participate to the study and fully fit the inclusion criteria. Thirty-seven (74%) among this population were female, and thirteen (26%) were male. The average age was 31 ± 11 years. Fifteen individuals aged between 21 and 52 years were healthy, and 35 patients aged between 18 and 62 years were identified as having periodontitis; however, there was no statistically significant difference between the two groups. Demographic and clinical characteristics of the study population are summarized in Table 1.

In the periodontitis group, subgroups were identified regarding staging and grade. Thus, the patients were diagnosed as having periodontitis stage II or III and grade B or C. The statistical analysis showed no significant difference between periodontitis stage II and III regarding the average age. However, age was lower in patients who had grade C periodontitis in comparison with those who had grade B, and the difference was statistically significant (Tables 2 and 3).

The growth of yeast colonies was recorded as a positive growth and the subject as a positive carrier. Among the 17 positive cultures, height isolates were identified as *C. albicans* by PCR (one healthy subject and 7 periodontitis patients) (Figure 3).

The presence of *C. albicans* was studied among the periodontitis population according to periodontal status. No significant differences were found between different groups (Table 4).

Analysis of the presence of *Candida albicans* between subgroups in the periodontitis group did not show any statistical difference neither when comparing stages II and III (33.3% and 17.2%, respectively, p = 0.370) nor when comparing grades Band C (21.0% and 18.7%, respectively, p = 0.865).

4. Discussion

Microbial-flora-associated periodontitis is very complex and has been continuously studied for many years. However, it should be emphasized that even if bacteria are considered as a primary agent in periodontal pathogenesis, other microorganisms, e.g., viruses and yeasts, are increasingly involved in the microbial etiology of periodontal diseases.

Candida albicans, the most widespread yeast in the oral cavity, can be isolated in healthy subjects without any clinical manifestation. It colonizes commonly the tongue [1]. However, many studies reported its presence in the healthy periodontium too. In the present study, 7% of the periodontal healthy patients harbored *C. albicans* in the subgingival plaque. These results are high in comparison to those reported by Urzúa et al. [21], which reported a prevalence of 3.57% in a population of 28 healthy subjects [21]. Meanwhile, Canabaro et al. [22] reported that 14.28% of 20 examined healthy subjects were yeast positive. Other studies showed variable occurrence (16% to 36%) of C. albicans in the subgingival plaque of the healthy periodontium [11, 23-25]. In immunocompetent subjects, C. albicans exists as a minor component of the oral biofilm [26]. It has been suggested also that its presence in the subgingival area could be transient [27]. However, it can also exist in periodontal pockets, and its role in periodontal pathogenesis is not vet clear.

In the present study, 20% of periodontitis patients were positive for *C. albicans*. Almost the same result had been

TABLE 1: Demographic and clinical characteristics of the study population	Table 1: Den	nographic and	clinical	characteristics	of the	study	population
---	--------------	---------------	----------	-----------------	--------	-------	------------

	Healthy $(n = 15; 30\%)$	Periodontitis ($n = 35; 70\%$)	Total $(n = 50; 100\%)$	p value
Gender (male/female)	5/10	8/27	13/37	p = 0.439
Age (years) (mean ± SD)	28.73 ± 9.24	33.31 ± 12.16	31.94 ± 11.47	p = 0.199
Plaque index (mean ± SD)	0.21 ± 0.10	2.41 ± 0.69	1.75 ± 1.17	$p \le 0.001$
Gingival index (mean ± SD)	0.05 ± 0.04	2.48 ± 0.68	1.75 ± 1.26	$p \le 0.001$
Pocket depth (mean ± SD)	2.80 ± 0.35	5.79 ± 1.83	4.90 ± 2.07	$p \le 0.001$
Clinical attachment loss (mean ± SD)	_	4.61 ± 1.24	_	_

Table 2: Comparison of clinical parameters in the periodontitis group according to stages II and stage III.

	Periodontitis group $(n = 35)$		
	Stage II $(n = 6; 17\%)$	Stage III $(n = 29; 83\%)$	p value
Age (mean ± SD)	33.00 ± 7.23	33.38 ± 13.04	0.946
Periodontal pockets depths (PPD) (mean ± SD)	5.00 ± 0.44	5.96 ± 1.97	0.251
Clinical attachment loss (CAL) (mean ± SD)	3.17 ± 0.2	4.91 ± 1.15	0.001

TABLE 3: Comparison of clinical parameters in the periodontitis group according to grade B and grade C.

	Periodontitis group $(n=35)$			
	Grade B $(n = 19; 54\%)$	Grade C $(n = 16; 46\%)$	p value	
Age (years) (mean \pm SD)	41.89 ± 8.73	23.13 ± 6.31	0.000	
Periodontal pockets depths (PPD) (mm) (mean ± SD)	4.78 ± 1.39	7.00 ± 1.56	0.000	
Clinical attachment loss (CAL) (mm) (mean ± SD)	4.12 ± 0.73	5.20 ± 1.47	0.008	

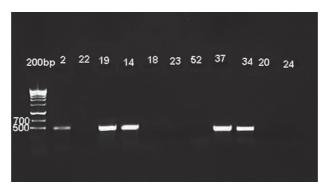


Figure 3: Agarose gel electrophoresis of PCR products of Candida albicans isolates.

Table 4: The presence of Candida albicans among the study population according to periodontal status.

		Candida	61	
		Negative carriers $(n = 28; 80\%)$	Positive carriers $(n = 7; 20\%)$	p value
Gender	Female Male	21 (78%) 07 (87.5%)	06 (22%) 01 (12.5%)	0.484
Age (years) (r	mean ± SD)	32.02 12.19	31.50 7.07	0.907
Plaque index (mean ± SD)	(mm)	2.37 ± 0.75	2.59 ± 0.34	0.460
Gingival index (mean ± SD)	x (mm)	2.47 ± 0.74	2.51 ± 0.39	0.907
Periodontal p (PPD) (mm)		6.07 ± 1.87	4.68 ± 1.20	0.072
Clinical attach (mm) (mean		4.81 ± 1.29	3.82 ± 0.55	0.059

found by Dahlén [28], who reported a prevalence of 17% and suggested that this yeast was more common in periodontitis patients than in healthy people. Nevertheless, no significant differences were found between these two groups (p > 0.05) in the present study.

Relatively few studies have analyzed the possible role of *Candida albicans* in periodontitis [23, 24, 27]. Matić Petrović et al. [25] reported that there is no impact of periodontal pocket depth on the presence of subgingival yeast. However, it should be pointed out that the mean probing depth in their study was 2.89 ± 0.944 for periodontitis patients compared to 2.02 ± 0.524 as the mean probing depth in healthy periodontium subjects.

Many studies showed an association between subgingival colonization by yeast subspecies, including Candida albicans, and the presence of severe periodontitis [6, 28]. Jarvensivu et al. [29] showed a prevalence of 16% in a population of 25 patients with chronic periodontitis. C. albicans remains the most common fungal pathogen found in patients with periodontitis [21, 22]. In the present study and according to the new classification of periodontal diseases and conditions, the periodontitis group included 2 distinguished subgroups based onstage case definition: stage II and stage III. No statistical difference was seen between stage II and stage III, regarding neither age nor the presence of Candida albicans. When comparing the periodontitis group based on grade, two subgroups were distinguished, a grade B group and grade C group. A significant difference was found for the average age between the two groups. Indeed, patients with grade C periodontitis were younger, and the estimate rate of periodontitis progression was rapid in comparison with group B periodontitis patients. However, there was no significant difference between these grade subgroups when considering the presence of Candida albicans. Thus, the present study suggests that the presence of Candida albicans in the periodontitis group seems not to be related to the stage or the grade. Nonetheless, these results must be considered with caution because of the small size of the study population. Moreover, all included patients were subjects seeking treatment in a hospital structure. Thus, further well-designed study, including a bigger size population, is needed to assess and understand the real contribution of Candida albicans in the modified microbiome associated with periodontitis.

The role of *C. albicans* in periodontitis pathogenesis is yet unclear. Indeed, this yeast could alter the oral microbiome and, therefore, influence significantly bacteria colonization [30, 31]. Coadherence between *Candida albicans* and some bacteria may help the formation of complex biofilms with mixed species affecting, therefore, the microbial pathogenesis; in [26, 32, 33], it was suggested also that *C. albicans* promotes bacterial invasion of host cells by anaerobic bacteria such as *P. gingivalis* and, thus, induces infections by anaerobic bacterial diseases.

In addition, *C. albicans* can grow either aerobically or anaerobically [34], which may explain their presence in deep periodontal pockets [22, 35]. Other studies reported that *C. albicans* is capable of adhering to epithelial cells and

induce inflammation [36]. *C. albicans* may have some virulence factors such as aspartyl proteinase (SAPs), phospholipases, and exoenzymes which disturb the locally immune response by the inhibition of polymorphonuclear neutrophil phagotocytosis and induce indirectly inflammatory reactions [24, 36, 37].

When comparing the presence of *C. albicans* regarding gender, 19% and 8%, respectively, of women and men were yeast positive. These results are the same as those reported by Reynaud et al. [24] who showed a prevalence of 20.3% among women compared to 8.2% in men. Nevertheless, as shown previously by Canabaro et al. [22], the presence of *Candida albicans* is not gender related (p = 0.342).

5. Conclusions

Candida albicans was present in periodontitis patients in this study. However, no statistical differences were found between the studied groups. Even if the role of *C. albicans* in periodontal disease has not yet been established, this yeast is considered an important pathogen in the progression and persistence of this disease. Thus, it would be interesting to study in depth the virulence factors and carry out in vitro studies to better understand the potential role of this yeast in this pathology and, especially, in refractory periodontal diseases.

Data Availability

All data used to support the findings of the study are included in the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

This work was supported by "Centre National pour la Recherche Scientifique et Technique, Morocco (CNRST)," under Grant no. PPR16/2016. The funder had no role in the study's design, data collection and analysis, the decision to publish, or the preparation of the manuscript.

References

- [1] T. M. Arendorf and D. M. Walker, "The prevalence and intraoral distribution of Candida albicans in man," *Archives of Oral Biology*, vol. 25, no. 1, pp. 1–10, 1980.
- [2] J. Kim and P. Sudbery, "Candida albicans, a major human fungal pathogen," *The Journal of Microbiology*, vol. 49, no. 2, pp. 171–177, 2011.
- [3] E. Hajishengallis, Y. Parsaei, M. I. Klein, and H. Koo, "Advances in the microbial etiology and pathogenesis of early childhood caries," *Molecular Oral Microbiology*, vol. 32, no. 1, pp. 24–34, 2017.
- [4] R. J. Lamont, H. Koo, and G. Hajishengallis, "The oral microbiota: dynamic communities and host interactions," *Nature Reviews Microbiology*, vol. 16, no. 12, pp. 745–759, 2018.

- [5] N. Buduneli and D. F. Kinane, "Host-derived diagnostic markers related to soft tissue destruction and bone degradation in periodontitis," *Journal of Clinical Periodontology*, vol. 38, pp. 85–105, 2011.
- [6] J. Slots, T. E. Rams, and M. A. Listgarten, "Yeasts, enteric rods and pseudomonads in the subgingival flora of severe adult periodontitis," *Oral Microbiology and Immunology*, vol. 3, no. 2, pp. 47–52, 1988.
- [7] A. Mombelli, "Microbiology and antimicrobial therapy of peri-implantitis," *Periodontology 2000*, vol. 28, pp. 177–189, 2002
- [8] R. Teles, F. Teles, J. Frias-Lopez, B. Paster, and A. Haffajee, "Lessons learned and unlearned in periodontal microbiology," *Periodontology 2000*, vol. 62, pp. 95–162, 2013.
- [9] C. Chen, P. Feng, and J. Slots, "Herpesvirus-bacteria synergistic interaction in periodontitis," *Periodontology 2000*, vol. 82, pp. 42–64, 2020.
- [10] K. Imai and Y. Ogata, "How does epstein-barr virus contribute to chronic periodontitis?" *International Journal of Molecular Sciences*, vol. 21, no. 6, p. 1940, 2020.
- [11] J. De-La-Torre, G. Quindós, C. Marcos-Arias et al., "Oral Candida colonization in patients with chronic periodontitis. Is there any relationship?" *Revista Iberoamericana de Micología*, vol. 35, no. 3, pp. 134–139, 2018.
- [12] D. Bartnicka, M. Gonzalez-Gonzalez, J. Sykut et al., "Candida albicans shields the periodontal killer porphyromonas gingivalis from recognition by the host immune system and supports the bacterial infection of gingival tissue," *Interna*tional Journal of Molecular Sciences, vol. 21, no. 6, Article ID 1984, 2020.
- [13] D. C. Coleman, D. J. Sullivan, D. E. Bennett, G. P. Moran, H. J. Barry, and D. B. Shanley, "Candidiasis," *Aids*, vol. 11, no. 5, pp. 557–567, 1997.
- [14] M. A. M. de Oliveira, L. P. Carvalho, M. D. S. Gomes, O. Bacellar, T. F. Barros, and E. M. Carvalho, "Microbiological and immunological features of oral candidiasis," *Microbiology* and *Immunology*, vol. 51, no. 8, pp. 713–719, 2007.
- [15] H. Löe, "The gingival index, the plaque index and the retention index systems," *Journal of Periodontology*, vol. 38, no. 6 Part II, pp. 610–616, 1967.
- [16] P. N. Papapanou, M. Sanz, N. Buduneli et al., "Periodontitis: consensus report of workgroup 2 of the 2017 world workshop on the classification of periodontal and peri-implant diseases and conditions," *Journal of Periodontology*, vol. 89, pp. S173–S182, 2018.
- [17] P. S. Bhavan, R. Rajkumar, S. Radhakrishnan, C. Seenivasan, and S. Kannan, "Culture and identification of Candida albicans from vaginal ulcer and separation of enolase on SDS-PAGE," *International Journal of Biological Macromolecules*, vol. 2, pp. 84–93, 2010.
- [18] J. C. O. Sardi, C. Duque, J. F. Höfling, and R. B. Gonçalves, "Genetic and phenotypic evaluation of Candida albicansstrains isolated from subgingival biofilm of diabetic patients with chronic periodontitis," *Medical Mycology*, vol. 50, no. 5, pp. 467–475, 2012.
- [19] R. Lamichhane, K. Boaz, S. Natarajan, and M. Shrestha, "Assessment of candidal carriage in patients with type II diabetes mellitus," *Journal of Pathology of Nepal*, vol. 5, no. 9, pp. 733–738, 2015.
- [20] M. Whiteway and C. Bachewich, "Morphogenesis inCandida albicans," *Annual Review of Microbiology*, vol. 61, no. 1, pp. 529–553, 2007.
- [21] B. Urzúa, G. Hermosilla, J. Gamonal et al., "Yeast diversity in the oral microbiota of subjects with periodontitis: Candida

- albicansandCandida dubliniensiscolonize the periodontal pockets," *Medical Mycology*, vol. 46, no. 8, pp. 783–793, 2008.
- [22] A. Canabarro, C. Valle, M. R. Farias, F. B. Santos, M. Lazera, and B. Wanke, "Association of subgingival colonization of Candida albicans and other yeasts with severity of chronic periodontitis," *Journal of Periodontal Research*, vol. 48, no. 4, pp. 428–432, 2013.
- [23] S. González, I. Lobos, A. Guajardo et al., "Yeasts in juvenile periodontitis," *Journal of Periodontology*, vol. 58, no. 2, pp. 119–124, 1987.
- [24] A. H. Reynaud, B. Nygaard-Østby, G.-K. Bøygard, E. R. Eribe, I. Olsen, and P. Gjermo, "Yeasts in periodontal pockets," *Journal of Clinical Periodontology*, vol. 28, no. 9, pp. 860–864, 2001.
- [25] S. Matić Petrović, M. Cimbaljević, M. Radunović, J. Kuzmanović Pfićer, A. Jotić, and A. Pucar, "Detection and sampling methods for isolation of Candidaspp. from oral cavities in diabetics and non-diabetics," *Brazilian Oral Re*search, vol. 29, pp. 1–7, 2015.
- [26] T. Wu, L. Cen, C. Kaplan et al., "Cellular components mediating coadherence of Candida albicans and fusobacterium nucleatum," *Journal of Dental Research*, vol. 94, no. 10, pp. 1432–1438, 2015.
- [27] T. E. Rams and J. Slots, "Candida biotypes in human adult periodontitis," *Oral Microbiology and Immunology*, vol. 6, no. 3, pp. 191-192, 1991.
- [28] G. Dahlén, "Role of suspected periodontopathogens in microbiological monitoring of periodontitis," *Advances in Dental Research*, vol. 7, no. 2, pp. 163–174, 1993.
- [29] A. Jarvensivu, J. Hietanen, R. Rautemaa, T. Sorsa, and M. Richardson, "Candida yeasts in chronic periodontitis tissues and subgingival microbial biofilms in vivo," *Oral Diseases*, vol. 10, no. 2, pp. 106–112, 2004.
- [30] M. M. Janus, W. Crielaard, C. M. C. Volgenant, M. H. Van Der Veen, B. W. Brandt, and B. P. Krom, "Candida albicans alters the bacterial microbiome of early in vitro oral biofilms," *Journal of Oral Microbiology*, vol. 9, no. 1, Article ID 1270613, 2017.
- [31] D. Bartnicka, K.-K. Justyna, Z. Marcin et al., "Adhesive protein-mediated cross-talk between Candida albicans and Porphyromonas gingivalis in dual species biofilm protects the anaerobic bacterium in unfavorable oxic environment," *Scientific Reports*, vol. 9, p. 4376, 2019.
- [32] H. Xu, H. F. Jenkinson, and A. Dongari-Bagtzoglou, "Innocent until proven guilty: mechanisms and roles of S treptococcus- C andida interactions in oral health and disease," *Molecular Oral Microbiology*, vol. 29, no. 3, pp. 99–116, 2014.
- [33] R. Tamai, M. Sugamata, and Y. Kiyoura, "Candida albicans enhances invasion of human gingival epithelial cells and gingival fibroblasts by Porphyromonas gingivalis," *Microbial Pathogenesis*, vol. 51, no. 4, pp. 250–254, 2011.
- [34] R. Dumitru, J. M. Hornby, and K. W. Nickerson, "Defined anaerobic growth medium for studying Candida albicans basic biology and resistance to eight antifungal drugs," *Antimicrobial Agents and Chemotherapy*, vol. 48, no. 7, pp. 2350–2354, 2004.
- [35] A. P. Vieira Colombo, C. B. Magalhães, F. A. R. R. Hartenbach, R. Martins Do Souto, and C. Maciel Da Silva-Boghossian, "Periodontal-disease-associated biofilm: a reservoir for pathogens of medical importance," *Microbial Pathogenesis*, vol. 94, pp. 27–34, 2016.
- [36] B. Hube and J. Naglik, "Candida albicans proteinases: resolving the mystery of a gene family," *Microbiology*, vol. 147, no. 8, pp. 1997–2005, 2001.

[37] N. A. Rubio, S. Puia, S. Toranzo, and M. I. Brusca, "Invasión fúngica en tejido conectivo en pacientes con enfermedad gingivo-periodontal," *Revista Iberoamericana de Micología*, vol. 32, no. 1, pp. 20–24, 2015.