




# Periodontitis and systemic markers of neurodegeneration: A case-control study

Yago Leira<sup>1,2,3</sup>  | Álvaro Carballo<sup>2</sup> | Marco Orlandi<sup>1</sup> | José Manuel Aldrey<sup>4,5</sup> | Juan Manuel Pías-Peleteiro<sup>4,5</sup> | Federico Moreno<sup>1</sup> | Laura Vázquez-Vázquez<sup>4,5</sup> | Francisco Campos<sup>5</sup> | Francesco D'Aiuto<sup>1</sup>  | José Castillo<sup>5</sup> | Tomás Sobrino<sup>5</sup> | Juan Blanco<sup>2,3</sup> 

<sup>1</sup>Periodontology Unit, UCL Eastman Dental Institute and NIHR UCLH Biomedical Research Centre, University College London, London, UK

<sup>2</sup>Periodontology Unit, Faculty of Medicine and Odontology, University of Santiago de Compostela, Santiago de Compostela, Spain

<sup>3</sup>Medical-Surgical Dentistry (OMEQUI) Research Group, Health Research Institute of Santiago de Compostela (IDIS), Santiago de Compostela, Spain

<sup>4</sup>Dementia Unit, Department of Neurology, Clinical University Hospital, Santiago de Compostela, Spain

<sup>5</sup>Clinical Neurosciences Research Laboratory, Health Research Institute of Santiago de Compostela (IDIS), Santiago de Compostela, Spain

## Correspondence

Yago Leira, Periodontology Unit - UCL Eastman Dental Institute and Hospital, University College London, 256 Gray's Inn Road, London WC1X 8LD, UK.  
Email: y.leira@ucl.ac.uk

## Funding information

This study was partially supported by grants from the Spanish Ministry of Economy and Competitiveness-Institute of Health Carlos III (PI13/02027 and PI15/01578), Spanish Ministry of Economy (RTI2018-102165-B-I00) and European Commission under the PANA project (Call H2020-NMP-2015-two stage, Grant 686009). YL holds a Senior Clinical Research Fellowship supported by the UCL Biomedical Research Centre who receives funding from the NIHR. Furthermore, FC (CP14/00154) and TS (CPII17/00027) are recipients of a research contract from Miguel Servet Programme of Institute of Health Carlos III. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## Abstract

**Aim:** To investigate whether periodontitis is associated with amyloid beta (A $\beta$ ) peptides and whether systemic inflammation could act as a potential mediator of this link.

**Materials and Methods:** A case-control study was designed including 75 patients with periodontitis (cases) and 75 age-balanced and gender-matched participants without periodontitis (controls). Full-mouth periodontal evaluation was performed in all participants. Demographic, clinical and behaviour data were also recorded. Fasting blood samples were collected, and serum levels of interleukin 6 (IL-6), high-sensitivity C-reactive protein (hs-CRP), A $\beta_{1-40}$  and A $\beta_{1-42}$  were determined.

**Results:** Cases showed higher levels of IL-6 ( $8.7 \pm 3.2$  vs.  $4.8 \pm 0.5$  pg/ml), hs-CRP ( $3.3 \pm 1.2$  vs.  $0.9 \pm 0.7$  mg/L), A $\beta_{1-40}$  ( $37.3 \pm 6.0$  vs.  $30.3 \pm 1.8$  pg/ml) and A $\beta_{1-42}$  ( $54.5 \pm 10.6$  vs.  $36.5 \pm 10.0$  pg/ml) when compared to controls (all  $p < .001$ ). Diagnosis of periodontitis was statistically significantly associated with circulating A $\beta_{1-40}$  ( $\beta$  coefficient<sub>adjusted</sub> = 6.9, 95% CI: 5.4–8.3;  $p < .001$ ) and A $\beta_{1-42}$  ( $\beta$  coefficient<sub>adjusted</sub> = 17.8, 95% CI: 14.4–21.3;  $p < .001$ ). Mediation analysis confirmed hs-CRP and IL-6 as mediators of this association.

**Conclusions:** Periodontitis is associated with increased peripheral levels of A $\beta$ . This finding could be explained by enhanced systemic inflammation that can be seen in patients with periodontitis.

## KEYWORDS

amyloid beta peptides, C-reactive protein, neurodegeneration, periodontitis, systemic inflammation

Sobrino and Blanco contributed equally as joint senior authors.

© 2020 John Wiley & Sons A/S. Published by John Wiley & Sons Ltd

## 1 | INTRODUCTION

Amyloid plaques contain small peptides of different lengths (from 39 to 43 amino acids), the so-called A $\beta$  peptides, which are the result of sequential proteolytic cleavage of the amyloid precursor protein (Murphy & LeVine, 2010). There are two main types of A $\beta$  peptides that are A $\beta$ <sub>1-40</sub> and A $\beta$ <sub>1-42</sub>. While A $\beta$ <sub>1-40</sub> is abundant and less neurotoxic, A $\beta$ <sub>1-42</sub> is less abundant and severely neurotoxic as well as more prone to aggregate to amyloid plaques (Tiwari, Atluri, Kaushik, & Yndart, 2019). Extracellular deposits of these A $\beta$  peptides in brain tissue are considered as one of the key hallmarks of Alzheimer's disease (AD), which is the most common form of age-related neurodegenerative disease (Magalingam, Radhakrishnan, Ping, & Haleagrahara, 2018). It has been shown that high levels of chronic inflammation measured by, for instance, acute-phase reactants [i.e. C-reactive protein (CRP)] or pro-inflammatory cytokines such as interleukin 6 (IL-6) can be toxic or further stimulate A $\beta$  production, aggregation and toxicity in the brain (Perry, 2004).

In the last decades, human periodontitis and its ultimate sequel (i.e. tooth loss) have been associated with cognitive decline/impairment in patients with AD (Holmer, Eriksdotter, Schultzberg, Pussinen, & Buhlin, 2018; Ide et al., 2016; Takeuchi et al., 2017). A recent meta-analysis of observational studies showed that subjects diagnosed with periodontitis have a 1.6-fold increased risk of developing AD (Leira et al., 2017). We recently demonstrated that bacterial endotoxin (i.e. lipopolysaccharide) from *Porphyromonas gingivalis* (*P. gingivalis*), the keystone pathogen of periodontitis, is capable of inducing systemic increase of A $\beta$  peptides (i.e. A $\beta$ <sub>1-40</sub> and A $\beta$ <sub>1-42</sub>) and this elevation correlated with alveolar bone loss in an animal model (Leira, Iglesias-Rey, et al., 2019). *P. gingivalis* and its toxic products can colonize areas from the brain resulting in enhanced neuroinflammation and overproduction of A $\beta$ <sub>1-42</sub>, which is part of amyloid plaques present in AD patients (Dominy et al., 2019). Tissues from otherwise healthy individuals but suffering from periodontitis showed overexpression of amyloid precursor protein (AAP), which produces A $\beta$  peptides (Kubota et al., 2014). Periodontitis was also associated with high A $\beta$ <sub>1-42</sub> concentrations in individuals with cognitive decline (Gil-Montoya et al., 2017). On the other hand, high levels of periodontal inflammation were recently related to systemic increase of A $\beta$ <sub>1-40</sub> in patients with a subtype of cerebral small vessel disease that is closely linked to vascular dementia and this association was mediated by enhanced systemic inflammation due to periodontitis (Leira, Rodríguez-Yáñez, et al., 2019a).

IL-6 is responsible for the regulation of the acute-phase response produced by the host after infection. CRP is one of the main acute-phase reactants and is primarily synthesized by IL-6 on the liver (Sproston & Ashworth, 2018). During inflammation, there is a linear correlation between increasing levels of IL-6 and CRP (Sproston & Ashworth, 2018). Therefore, both molecules are widely used to assess the presence and severity of low-grade inflammation (Sproston & Ashworth, 2018). Patients with untreated periodontitis show increased of circulating levels of CRP or IL-6, which reflects higher levels of inflammation beyond the oral cavity in this population (Loos, Craandijk, Hoek, Wertheim-van Dillen, & Velden, 2000).

### Clinical Relevance

*Scientific rationale for the study:* Peripheral inflammation is associated with high load of A $\beta$  peptides in Alzheimer's disease (AD). Periodontitis increases the risk of having cognitive decline; however, little is known on the potential role of periodontitis as a contributor to increased systemic concentrations of A $\beta$  peptides in otherwise healthy individuals. *Principal findings:* Circulating A $\beta$  peptides are elevated in periodontitis. A link between diagnosis of periodontitis and A $\beta$  was observed, and enhanced systemic inflammation could explain this relationship.

*Practical implications:* Our results suggest that periodontitis via systemic inflammation could contribute to increased peripheral levels of A $\beta$  peptides, which are key elements in the pathogenesis of AD.

Nevertheless, to the best of our knowledge, no human studies have investigated the effect of periodontitis on peripheral A $\beta$  peptides in systemically healthy individuals. Our hypothesis was that patients with periodontitis have higher serum levels of A $\beta$ <sub>1-40</sub> and A $\beta$ <sub>1-42</sub> and that systemic inflammation could mediate this increase. Hence, the aim of the study was twofold. The primary objective was to investigate whether there is an association between diagnosis of periodontitis and A $\beta$  peptides. As a secondary objective, we tested whether systemic inflammation could be a mediator of this relationship.

## 2 | MATERIALS AND METHODS

### 2.1 | Study design and participants

For this age-balanced and gender-matched case-control study, 150 participants otherwise healthy were recruited from the Faculty of Odontology of Santiago de Compostela (Spain) during the period comprised between January 2014 and April 2017. Cases consisted of 75 patients diagnosed with periodontitis (Eke, Page, Wei, Thornton-Evans, & Genco, 2012) among referrals to the Periodontology Unit (University of Santiago de Compostela, Spain) for diagnosis and treatment of periodontitis. Seventy-five controls were defined as those without any clinical/radiographic signs of periodontal disease (including gingivitis and periodontitis) and/or history of this disease. These participants were identified among friends of the patients with periodontitis ( $n = 29$ ) or from a research registry of participants from previous studies carried out by our research group ( $n = 46$ ) (Ameijeira, Leira, Domínguez, Leira, & Blanco, 2019; Leira & Blanco, 2018; Leira, Rodríguez-Yáñez, et al., 2019a).

Exclusion criteria were as follows: (a) <18 years of age; (b) <15 teeth (excluding third molars and retained roots); (c) periodontal treatment in the last year; (d) evident neurological diseases

confirmed clinically and/or by computed tomography/magnetic resonance imaging (e.g. neuroinflammatory, neurovascular or neurodegenerative conditions); (e) concomitant medical conditions (e.g. diabetes, cardiovascular diseases, hypertension or hypercholesterolemia) or active infectious/inflammatory diseases (e.g. HIV, hepatitis, tuberculosis, rheumatoid arthritis, allergies or asthma); (f) pregnant or lactating females; (g) malignancy; and (h) treatment with systemic antibiotics, corticosteroids and/or immunosuppressive agents within 3 months prior to periodontal examination (Figure S1).

This research was performed in accordance with the Declaration of Helsinki of the World Medical Association (2008) and approved by the Ethics Committee of the Serviço Galego de Saúde (ID: 2016/399). Written informed consent was obtained from each participant or their relatives after full explanation of the periodontal examination and blood sample collection. The study was performed following the STROBE (Strengthening the Reporting of Observational Studies in Epidemiology) guidelines (Von Elm et al., 2008).

## 2.2 | Periodontal assessment and demographic information

A full-mouth periodontal examination was performed in all participants by a calibrated periodontist (YL) (Ameijeira et al., 2019; Leira & Blanco, 2018; Leira, Rodríguez-Yáñez, et al., 2019a). The following parameters were measured in all teeth (except third molars): probing pocket depth (PPD), clinical attachment level (CAL), full-mouth plaque score (FMPS) and full-mouth bleeding score (FMBS) (Ainamo & Bay, 1975). Measurements were recorded at six sites per tooth (mesiobuccal, distobuccal, midbuccal, mesiolingual, distolingual and midlingual) using a calibrated University of North Carolina periodontal probe (UNC 15; Hu-Friedy). Slight periodontitis was defined as  $\geq 2$  inter-proximal sites with CAL  $\geq 3$  mm and  $\geq 2$  inter-proximal sites with PPD  $\geq 4$  mm (not on the same tooth) or one site with PPD  $\geq 5$  mm. Moderate periodontitis was defined as  $\geq 2$  inter-proximal sites with CAL of  $\geq 4$  mm (not on the same tooth) or  $\geq 2$  inter-proximal sites with PPD of  $\geq 5$  mm, also not on the same tooth. Severe periodontitis was defined as the presence of  $\geq 2$  inter-proximal sites with CAL of  $\geq 6$  mm (not on the same tooth) and  $\geq 1$  inter-proximal sites with PPD of  $\geq 5$  mm. Total periodontitis was the sum of slight, moderate and severe periodontitis (Eke et al., 2012).

In addition, the periodontal inflamed surface area (PISA) was calculated for all participants. PISA reflects the surface area of bleeding pocket epithelium in  $\text{mm}^2$ . As previously described (Leira & Blanco, 2018; Leira, Rodríguez-Yáñez, et al., 2019a, Leira, Rodríguez-Yáñez, et al., 2019b), PISA was calculated using a Microsoft Excel spreadsheet, in the following steps: (a) mean CAL and gingival recession for each particular tooth are calculated; (b) linear mean CAL and gingival recession are translated into the periodontal epithelial surface area (PESA) for each specific tooth (Hujuel, White, García, & Listgarten, 2001). The PESA for a particular tooth consists of the root surface area of that tooth measured in  $\text{mm}^2$ , which

is covered with pocket epithelium; (c) the PESA for a specific tooth is then multiplied by the proportion of sites around the tooth that was affected by bleeding on probing, resulting in the PISA for that particular tooth; and (d) the sum of all individual PISAs around individual teeth is calculated, rendering the full-mouth PISA value in  $\text{mm}^2$  for each participant (Nesse et al., 2008).

For both groups, smoking status was evaluated by a questionnaire. Body mass index (BMI) was calculated for all participants using the formula  $\text{weight}/\text{height}^2$  ( $\text{kg}/\text{m}^2$ ).

## 2.3 | Sample collection and laboratory tests

Fasting blood samples were obtained in the morning at the same time as the periodontal assessment and interview. Briefly, 2 ml of venous blood was collected from the antecubital fossa by venepuncture using a 20-gauge needle with a 2-ml syringe. Blood samples were allowed to clot at room temperature and, after 1 hr, serum was separated by centrifugation (15 min at 3,000 g) and 0.5 ml of extracted serum was immediately transferred to 1.5-ml aliquots. Each aliquot was stored at  $-80^\circ\text{C}$  until required for analysis. Serum levels of high-sensitive CRP (hs-CRP) were measured using an immunodiagnostic IMMULITE<sup>®</sup> 2000 Systems (Siemens Healthcare Diagnostics); minimum assay sensitivity was 0.2 mg/L. Serum levels of IL-6 and A $\beta$  peptides were measured by enzyme-linked immunosorbent assay (ELISA) technique following the manufacturer's instructions IL-6 ELISA kit (Proteintech<sup>™</sup>); minimum assay sensitivity was 3.8 pg/ml; A $\beta_{1-40}$  ELISA kit (Elabscience<sup>®</sup>); minimum assay sensitivity was 9.38 pg/ml with an intra-assay coefficient of variation (CV) of 4.6% and inter-assay CV of 6.5%, and A $\beta_{1-42}$  ELISA kit (Elabscience<sup>®</sup>); and minimum assay sensitivity was 9.38 pg/ml with an intra-assay CV of 6.0% and inter-assay CV of 6.8%.

Determinations were performed in an independent laboratory blinded to clinical data. Clinical investigators were unaware of the laboratory results until the study had ended. hs-CRP was determined in the Central Laboratory of the Clinical University Hospital of Santiago de Compostela, whereas A $\beta$  peptides and IL-6 were determined in the Clinical Neurosciences Research Laboratory of the same hospital.

## 2.4 | Statistical analysis

For this observational study, a sample size of 69 patients per group (case vs. controls) (1:1 ratio) was sufficient to detect a 5 pg/ml difference in serum A $\beta_{1-40}$  between study groups, with a standard deviation of 9 pg/ml and assuming  $\alpha$ -value = 0.05 and statistical power of 90% (Leira, Rodríguez-Yáñez, et al., 2019a).

Mean values  $\pm$  standard deviation (SD) and median [P<sub>25</sub>, P<sub>75</sub>] were calculated for normally and non-normally distributed continuous variables, respectively. Statistical tests used to compare continuous data were *independent t-test* or *Mann-Whitney U test*. Categorical

variables were reported as percentages and compared by chi-square test. Non-parametric correlation analyses between biomarkers and clinical periodontal parameters were performed using Spearman's rank correlation coefficient.

Linear regression models adjusted for confounders (i.e. age, gender, BMI and smoking) were created to test association between periodontitis and A $\beta$  peptides as well as with A $\beta_{42:40}$  ratio. Mediation analysis was carried out to investigate whether systemic inflammation (measured by hs-CRP and IL-6) could explain the potential association between periodontitis and A $\beta$  peptides as well as with A $\beta_{42:40}$  ratio. In order to do so, the PROCESS macro for SPSS (Hayes, 2013) was used. Each analytical step was adjusted for confounders previously named. Mediation analysis tests whether the association between a predictor/exposure (X = periodontitis) and an outcome (Y = A $\beta$  peptides and A $\beta_{42:40}$  ratio) is mediated through a mediator (M = systemic inflammatory markers). Estimates with 95% confidence intervals were calculated using ordinary least square regression for each step. The indirect effect of the predictor on the outcome mediated via the mediator (also named mediated effect) was considered significant if the corresponding 95% bootstrap confidence interval did not include zero. This approach is the one recommended by Hayes due to it does not assume a normal distribution of the sample (Hayes, 2013). The percentage of the effect of periodontitis on A $\beta$  peptides and A $\beta_{42:40}$  ratio mediated by markers of systemic inflammation was also calculated using the following formula: (mediated effect/total effect)\*100.

All tests were carried out at a significance level of  $\alpha = 0.05$  using IBM SPSS Statistics (version 24.0).

### 3 | RESULTS

#### 3.1 | Demographic, clinical and periodontal data

Table 1 depicts participant's characteristics and demographic factors. Cases and controls were balanced for age and matched for gender. No statistically significant differences were observed in terms of BMI and smoking status.

Cases had greater periodontal inflammation when compared to controls (Table 1). Both measures of active gingival inflammation (PPD, FMBS and PISA) and historic periodontal tissue attachment loss (CAL) were significantly higher in cases than in controls. Also, the level of bacterial plaque accumulation was greater in those with periodontitis than in controls. On average, cases had one fewer tooth present in the mouth compared to those without periodontitis (Table 1). Within periodontal patients, 23 (30.7%) had slight periodontitis, 30 (40.0%) presented moderate periodontitis and 22 (29.3%) had severe periodontitis.

#### 3.2 | Biomarkers

Cases presented statistically significantly higher serum levels of IL-6, hs-CRP, A $\beta_{1-40}$  and A $\beta_{1-42}$  compared to controls (all  $p < .001$ ; Table 2).

**TABLE 1** Baseline characteristics

Variables	Periodontitis (n = 75)	No periodontitis (n = 75)	p-value
Age (years)	44.8 $\pm$ 10.3	44.7 $\pm$ 12.2	.983
Gender, male, n (%)	49 (65.3)	49 (65.3)	1.000
BMI (kg/m <sup>2</sup> )	25.5 (12.9, 29.4)	23.4 (21.0, 27.4)	.079
Smoking status, n (%)			
Current	11 (14.7)	7 (9.3)	.252
Former	8 (10.7)	4 (5.3)	
Never	56 (74.7)	64 (85.3)	
Periodontal clinical parameters			
PPD (mm)	3.2 $\pm$ 0.9	2.0 $\pm$ 0.4	<b>&lt;.001</b>
CAL (mm)	3.5 $\pm$ 1.0	2.1 $\pm$ 0.5	<b>&lt;.001</b>
FMBS (%)	57.8 $\pm$ 23.4	23.8 $\pm$ 8.8	<b>&lt;.001</b>
FMPS (%)	66.2 $\pm$ 17.3	27.9 $\pm$ 12.1	<b>&lt;.001</b>
PISA (mm <sup>2</sup> )	610.8 (320.9, 1,139.0)	26.5 (17.5, 43.8)	<b>&lt;.001</b>
Teeth present, n	26.2 $\pm$ 1.8	27.0 $\pm$ 0.7	<b>.017</b>

Significant results are reported in bold.

Abbreviations: BMI, body mass index; CAL, clinical attachment level; FMBS, full-mouth bleeding score; FMPS, full-mouth plaque score; PISA, periodontal inflamed surface area; PPD, probing pocket depth.

Serum levels of IL-6 (12.4  $\pm$  3.6 vs. 7.9  $\pm$  0.9 pg/ml,  $p < .001$  and vs. 6.3  $\pm$  0.8 pg/ml,  $p < .001$ ; Figure 1a), hs-CRP (4.9  $\pm$  0.9 vs. 3.1  $\pm$  0.2 mg/L,  $p < .001$  and vs. 2.2  $\pm$  0.3,  $p < .001$ ; Figure 1b), A $\beta_{1-40}$  (43.9  $\pm$  4.9 vs. 36.1  $\pm$  4.0 pg/ml,  $p < .001$  and vs. 32.6  $\pm$  2.9 pg/ml,  $p < .001$ ; Figure 1c) and A $\beta_{1-42}$  (66.2  $\pm$  6.1 vs. 54.5  $\pm$  5.4 pg/ml,  $p < .001$  and vs. 43.3  $\pm$  5.8 pg/ml,  $p < .001$ ; Figure 1d) were statistically significantly elevated in severe periodontal patients in comparison with those cases with moderate or slight forms of periodontitis. Similarly, the moderate group presented higher levels of IL-6 (7.9  $\pm$  0.9 vs. 6.3  $\pm$  0.8 pg/ml,  $p = .001$ ; Figure 1a), hs-CRP (3.1  $\pm$  0.2 mg/L vs. 2.2  $\pm$  0.3,  $p < .001$ ; Figure 1b), A $\beta_{1-40}$  (36.1  $\pm$  4.0 vs. 32.6  $\pm$  2.9 pg/ml,  $p = .015$ ; Figure 1c) and A $\beta_{1-42}$  (54.5  $\pm$  5.4 vs. 43.3  $\pm$  5.8 pg/ml,  $p = .005$ ; Figure 1d) than slight periodontal patients. When levels of each biomarkers were compared between each grade of severity of periodontitis and the group of participants with a healthy periodontium, statistically significant differences were also found [for IL-6: healthy (4.8  $\pm$  0.5 pg/ml) vs. (slight (6.3  $\pm$  0.8 pg/ml,  $p < .001$ ), moderate (7.9  $\pm$  0.9 pg/ml,  $p < .001$ ) and severe (12.4  $\pm$  3.6 pg/ml,  $p < .001$ ); Figure 1a); for hs-CRP: healthy (0.9  $\pm$  0.7 mg/L) vs. (slight (2.2  $\pm$  0.3,  $p < .001$ ), moderate (3.1  $\pm$  0.2 mg/L,  $p < .001$ ) and severe (4.9  $\pm$  0.9 mg/L,  $p < .001$ ); Figure 1b); for A $\beta_{1-40}$ : healthy (30.3  $\pm$  1.8 ng/ml) vs. (slight (32.6  $\pm$  2.9 pg/ml,  $p = .015$ ), moderate (36.1  $\pm$  4.0 pg/ml,  $p < .001$ ) and severe (43.9  $\pm$  4.9 pg/ml,  $p < .001$ ); Figure 1c); and for A $\beta_{1-42}$ : healthy (36.5  $\pm$  10.0 pg/ml) vs. (slight (43.3  $\pm$  5.8 pg/ml,  $p = .005$ ), moderate (54.5  $\pm$  5.4 pg/ml,  $p < .001$ ) and severe (66.2  $\pm$  6.1 pg/ml,  $p < .001$ ); Figure 1d)].

**TABLE 2** Biochemical parameters

Variables	Periodontitis (n = 75)	No periodontitis (n = 75)	p-value
Systemic inflammation			
IL-6 (pg/ml)	8.7 ± 3.2	4.8 ± 0.5	<b>&lt;.001</b>
hs-CRP (mg/L)	3.3 ± 1.2	0.9 ± 0.7	<b>&lt;.001</b>
Aβ peptides			
Aβ <sub>1-40</sub> (pg/ml)	37.3 ± 6.0	30.3 ± 1.8	<b>&lt;.001</b>
Aβ <sub>1-42</sub> (pg/ml)	54.5 ± 10.6	36.5 ± 10.0	<b>&lt;.001</b>
Aβ <sub>42:40</sub> (pg/ml)	1.5 ± 0.2	1.2 ± 0.3	<b>&lt;.001</b>

Significant results are reported in bold.

Abbreviations: Aβ<sub>1-40</sub> and Aβ<sub>1-42</sub>, amyloid beta 1-40 and 1-42; Aβ<sub>42:40</sub>, amyloid beta 1-40 and 1-42 ratio; hs-CRP, high-sensitivity C-reactive protein; IL-6, interleukin 6.

Aβ<sub>42:40</sub> ratio was higher in cases than controls (Table 2). Compared to periodontally healthy subjects (1.2 ± 0.3 pg/ml), greater Aβ<sub>42:40</sub> ratio was observed in severe (1.5 ± 0.1,  $p < .001$ ) and moderate cases (1.5 ± 0.2,  $p < .001$ ) but not for slight periodontal individuals (1.3 ± 0.2,  $p = .507$ ). No statistically significant differences were found between different grades of severity of periodontitis (data not shown).

### 3.3 | Correlation between biomarkers and clinical periodontal variables

Strong positive correlations were found between clinical periodontal parameters (PPD, CAL, FMBS, FMPS and PISA) and IL-6, hs-CRP, Aβ<sub>1-40</sub> and Aβ<sub>1-42</sub> as well as Aβ<sub>42:40</sub> ratio (Table 3). IL-6 correlated positively with Aβ<sub>140</sub> and Aβ<sub>1-42</sub> ( $r = .250$ ,  $p < .001$  and  $r = .735$ ,  $p < .001$ ; respectively; Figure 2a,b). Similarly, hs-CRP strongly correlated with both Aβ<sub>1-40</sub> ( $r = .656$ ,  $p < .001$ ) and Aβ<sub>1-42</sub> ( $r = .702$ ,  $p < .001$ ; Figure 2c,d). Statistically significant correlations were also found for Aβ<sub>42:40</sub> ratio (IL-6:  $r = .453$ ,  $p < .001$  and hs-CRP:  $r = .401$ ,  $p < .001$ ).

### 3.4 | Association between periodontitis and Aβ peptides and mediation analysis

Diagnosis of periodontitis was statistically significantly associated with circulating Aβ<sub>1-40</sub> ( $\beta$  coefficient<sub>adjusted</sub> = 6.9, 95% CI: 5.4–8.3;  $p < .001$ ) and Aβ<sub>1-42</sub> ( $\beta$  coefficient<sub>adjusted</sub> = 17.8, 95% CI: 14.4–21.3;  $p < .001$ ) as well as with Aβ<sub>42:40</sub> ratio ( $\beta$  coefficient<sub>adjusted</sub> = 0.2, 95% CI: 0.1–0.3;  $p < .001$ ).

Mediation analysis showed a statistically significant effect of both markers of systemic inflammation (hs-CRP and IL-6) mediating the association between periodontitis and both Aβ peptides (Table 4). Periodontitis was statistically significantly associated with increased levels of systemic inflammation (all  $p < .001$ ).

Systemic inflammatory markers were also statistically significantly related to higher levels of Aβ peptides (all  $p < .001$ ). The indirect (mediated) effect of periodontitis on Aβ peptides mediated through hs-CRP and IL-6 was statistically significant (95% CI did not include 0). The percentage mediated in the association between periodontitis and Aβ peptides through hs-CRP and IL-6 ranged from 38% to 82% (Table 4). Similar findings were observed when continuous measures of periodontitis were used as exposure, for example PPD and CAL (data not shown). No significant mediation effect was found when Aβ<sub>42:40</sub> ratio was used as an outcome (Table 4).

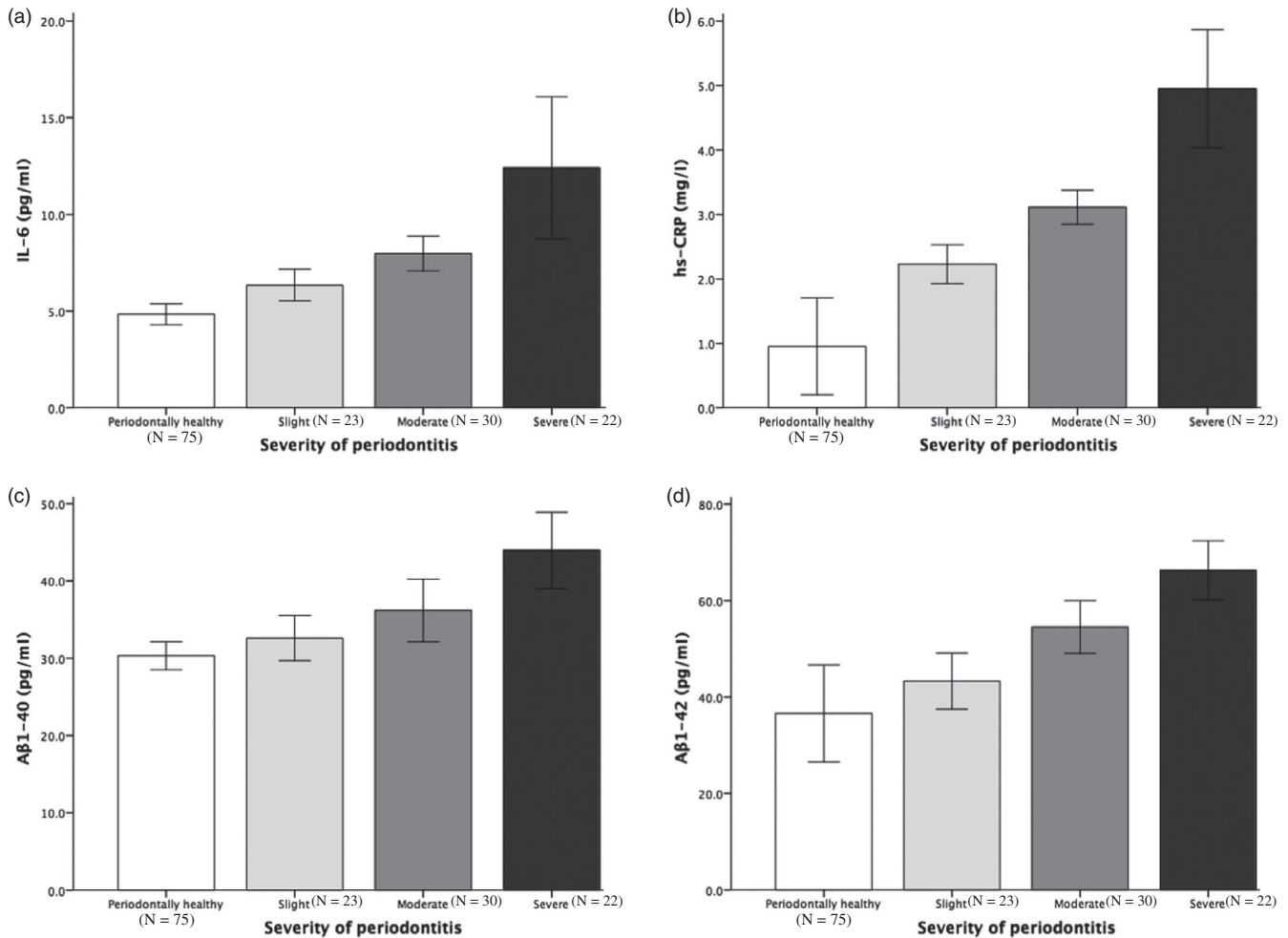
## 4 | DISCUSSION

In the present age-balanced and gender-matched case-control study, periodontitis was associated with increased peripheral levels of Aβ peptides and this relationship was mediated by systemic inflammation.

The link between periodontitis and systemic inflammation is well documented (Amar et al., 2003; Loos et al., 2000; Paraskevas, Huizinga, & Loos, 2008). Further, periodontal treatment reduces pro-inflammatory markers measured in peripheral blood (D'Aiuto et al., 2004). In the present study, we confirmed that periodontitis was associated with elevated serum levels of hs-CRP and IL-6 and this increase was linked to severity of periodontitis. Previous observational studies from different cohorts demonstrated increased level of systemic inflammation (measured by circulating CRP concentrations) in patients with periodontitis (Ardila & Guzmán, 2015; Linden, McClean, Young, Evans, & Kee, 2008; Noack et al., 2001; Pitiphat, Savetsilp, & Wara-Aswapati, 2008; Slade, Offenbacher, Beck, Heiss, & Pankow, 2000). In the present study, clinical parameters of current/active periodontal inflammation (PPD, FMBS and PISA) and history of periodontal attachment loss (CAL) were positively correlated with both markers of systemic inflammation.

Aβ deposits play an important role in the pathogenesis of AD, as they are the main component of senile plaques in brain grey matter (Murphy & LeVine, 2010). Extracellular aggregation of Aβ plaques is associated with widespread neuronal atrophy resulting in gradual neuronal death and memory loss. The most relevant finding in the present study is the overexpression of Aβ peptides (Aβ<sub>140</sub> and Aβ<sub>1-42</sub>) in periodontitis subjects with good general health. Systemic levels of Aβ peptides were significantly elevated in patients with periodontitis compared to controls. Also, clinical periodontal parameters positively correlated with increased Aβ peptide concentrations. Previously, it has been reported that in cognitively normal healthy elderly, measures of periodontitis such as CAL were associated with high brain amyloid accumulation assessed by positron emission tomography (Kamer et al., 2015). The present results are in line with a recent animal experiment carried out by our group, which has shown that bacterial endotoxin from *P. gingivalis* evoked a raise in peripheral Aβ peptides in systemically healthy rats during 21 days of follow-up (Leira, Iglesias-Rey,





**FIGURE 1** Serum levels of biomarkers according to severity of periodontitis [slight ( $N = 23$ ), moderate ( $N = 30$ ) and severe ( $N = 22$ ): (a) IL-6; (b) hs-CRP; (c) A $\beta_{1-40}$ ; (d) A $\beta_{1-42}$

	PPD (mm)	CAL (mm)	FMBS (%)	FMPS (%)	PISA (mm <sup>2</sup> )
IL-6 (pg/ml)	0.702	0.692	0.680	0.636	0.821
<i>p</i> -value	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>
hs-CRP (mg/L)	0.730	0.723	0.764	0.701	0.895
<i>p</i> -value	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>
A $\beta_{1-40}$ (pg/ml)	0.625	0.640	0.610	0.529	0.692
<i>p</i> -value	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>
A $\beta_{1-42}$ (pg/ml)	0.636	0.661	0.621	0.502	0.740
<i>p</i> -value	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>
A $\beta_{42:40}$ (pg/ml)	0.360	0.381	0.330	0.300	0.429
<i>p</i> -value	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>	<b>&lt;.001</b>

Significant results are reported in bold.

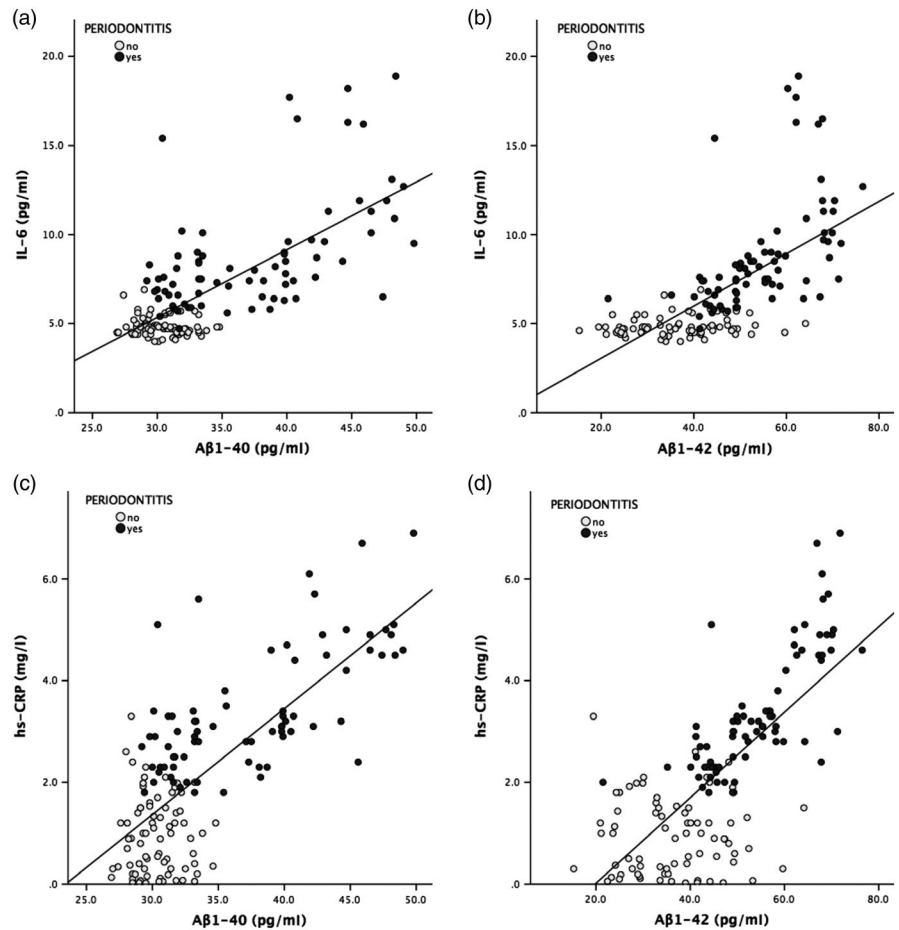
Abbreviations: A $\beta_{1-40}$  and A $\beta_{1-42}$ , amyloid beta 1-40 and 1-42; A $\beta_{42:40}$ , amyloid beta 1-40 and 1-42 ratio; CAL, clinical attachment level; FMBS, full-mouth bleeding score; FMPS, full-mouth plaque score; hs-CRP, high-sensitivity C-reactive protein; IL-6, interleukin 6; PISA, periodontal inflamed surface area; PPD, probing pocket depth.

**TABLE 3** Spearman's correlation coefficient for clinical periodontal parameters and biomarkers

et al., 2019). Other experiments have also demonstrated that chronic exposure of *P. gingivalis* and its toxins is capable of inducing brain colonization and induction of A $\beta_{1-42}$ , microglia-mediated

neuroinflammation as well as learning and memory impairment (Dominy et al., 2019; Wu et al., 2017), thus supporting the hypothesis that a chronic peripheral inflammatory condition such as

**FIGURE 2** Spearman's correlation coefficient for systemic inflammatory markers and A $\beta$  peptides: (a) IL-6 and A $\beta_{1-40}$ ; (b) IL-6 and A $\beta_{1-42}$ ; (c) hs-CRP and A $\beta_{1-40}$ ; (d) hs-CRP and A $\beta_{1-42}$



periodontitis could be involved in the onset/progression of AD. In addition to neurotoxicity, A $\beta_{1-40}$  can also contribute to cerebral vascular pathology and endothelial dysfunction when deposited in cerebral microvessels (Iadecola, Park, & Capone, 2009; Niwa, Carlson, & Iadecola, 2000; Niwa, Younkin, et al., 2000). Indeed, we recently showed that active periodontal inflammation was associated with increased systemic levels of A $\beta_{1-40}$  in patients diagnosed with a subtype of cerebral small vessel disease (lacunar infarcts) that could cause vascular dementia (Leira, Rodríguez-Yáñez, et al., 2019a).

Different mechanisms may explain why periodontitis subjects had higher circulating levels of A $\beta$  peptides. On the one hand, a direct production of A $\beta$  could occur in periodontitis. It has been shown that gingival tissues with periodontitis are able to express specific genes related to A $\beta$  production such as APP mRNA (Kubota et al., 2014). On the other hand, an indirect pathway by which A $\beta$  is produced due to periodontitis could be also speculated. Evidence suggests that systemic inflammation can produce A $\beta$  in the systemic circulation and promotes its accumulation in the brain (Perry, 2004). Our results showed a positive correlation between systemic inflammatory markers (hs-CRP and IL-6) and A $\beta$  peptides, therefore explaining the association between periodontitis, systemic inflammation and A $\beta$  overexpression (Wang, Ho, Leung, Goto, & Chang, 2019). In addition, mediation analysis

showed that markers of systemic inflammation (hs-CRP and IL-6) acted as mediators in the association between periodontitis and A $\beta$  peptides; therefore, increased systemic inflammation could be a plausible biological explanation of our findings. However, more research is needed in this area to elucidate the exact mechanisms behind this relationship because when the A $\beta_{42:40}$  ratio was used as an outcome no significant mediation effect was observed by inflammation perhaps because inflammatory markers were not associated with A $\beta_{42:40}$  ratio. For instance, whether systemic A $\beta$  peptides in patients with periodontitis are able to cross the blood-brain barrier and accumulate on the brain or whether systemic inflammatory markers can actually induce neuroinflammation in these patients and therefore lead to neuronal damage.

Some limitations of the present study should be addressed. Firstly, the retrospective design of the study does not allow us to test causality on the association between periodontitis and circulating A $\beta$  peptides. Secondly, participants were in good systemic health but this was confirmed by means of a self-reported questionnaire and some of them might presented with poor metabolic control (including high blood pressure, poor glycaemic control and dyslipidaemia) and undiagnosed hypertension, diabetes or hypercholesterolemia as well as high BMI which could have influenced levels of both systemic inflammation and A $\beta$  peptides (de Miguel, Rudemiller, Abais, & Mattson, 2015; Ellulu, Patimah, Khaza'ai, Rahmat, & Abed, 2017; Lee

et al., 2008; Marsland, McCaffery, Muldoon, & Manuck, 2010; Razay, Vreugdenhil, & Wilcock, 2007; Shah et al., 2012). Thirdly, some of the controls were friends of cases. This fact could have led to selection

bias as they could be more motivated and have higher response rates compared to general population controls that do not know the case. Also these people might be more sociable and extrovert as they are

**TABLE 4** Mediation of systemic inflammation for the association between periodontitis and A $\beta$  peptides as well as with A $\beta_{42:40}$  ratio

Model A (exposure: periodontitis/outcome: A $\beta_{1-40}$ )		Mediator: hs-CRP		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	2.40	0.17	<.001	2.07–2.74
b (mediator $\rightarrow$ outcome)	2.34	0.31	<.001	1.72–2.96
c (total effect)	6.86	0.75	<.001	5.38–8.34
c' (direct effect)	1.23	0.99	.22	–0.74 to 3.18
ab (mediated effect)	5.63	1.04	–	3.58–7.62
ab/c (hs-CRP percentage mediated) = 82%				
Model B (exposure: periodontitis/outcome: A $\beta_{1-40}$ )		Mediator: IL-6		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	3.81	0.38	<.001	3.05–4.56
b (mediator $\rightarrow$ outcome)	0.98	0.14	<.001	0.70–1.26
c (total effect)	6.86	0.75	<.001	5.38–8.34
c' (direct effect)	3.14	0.85	<.001	1.47–4.81
ab (mediated effect)	3.72	0.75	–	2.31–5.23
ab/c (IL-6 percentage mediated) = 54%				
Model C (exposure: periodontitis/outcome: A $\beta_{1-42}$ )		Mediator: hs-CRP		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	2.40	0.17	<.001	2.07–2.74
b (mediator $\rightarrow$ outcome)	4.56	0.76	<.001	3.05–6.07
c (total effect)	17.84	1.73	<.001	14.42–21.26
c' (direct effect)	6.87	2.41	<.001	2.12–11.63
ab (mediated effect)	10.97	2.09	–	6.79–14.95
ab/c (hs-CRP percentage mediated) = 61%				
Model D (exposure: periodontitis/outcome: A $\beta_{1-42}$ )		Mediator: IL-6		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	3.81	0.38	<.001	1.59–6.85
b (mediator $\rightarrow$ outcome)	1.80	0.35	<.001	1.11–2.48
c (total effect)	17.84	1.73	<.001	14.42–21.26
c' (direct effect)	11.00	2.07	<.001	6.92–15.09
ab (mediated effect)	6.84	1.36	–	4.56–9.76
ab/c (IL-6 percentage mediated) = 38%				
Model E (exposure: periodontitis/outcome: A $\beta_{42:40}$ )		Mediator: hs-CRP		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	2.40	0.16	<.001	2.06–2.73
b (mediator $\rightarrow$ outcome)	0.03	0.02	.21	–0.01 to 0.07
c (total effect)	0.25	0.05	<.001	0.35–0.78
c' (direct effect)	0.17	0.07	.02	0.02–0.33
ab (mediated effect)	0.07	0.05	–	–0.03 to 0.17

(Continues)



TABLE 4 (Continued)

Model E (exposure: periodontitis/outcome A $\beta_{42:40}$ )		Mediator: hs-CRP		
Effect	Estimate	SE	p-value	95% CI
ab/c (hs-CRP percentage mediated) = no mediation				
Model F (exposure: periodontitis/outcome A $\beta_{42:40}$ )		Mediator: IL-6		
Effect	Estimate	SE	p-value	95% CI
a (exposure $\rightarrow$ mediator)	3.80	0.38	<.001	3.04–4.56
b (mediator $\rightarrow$ outcome)	0.01	0.01	.30	–0.01 to 0.03
c (total effect)	0.25	0.05	<.001	0.35–0.78
c' (direct effect)	0.21	0.06	.001	0.08–0.33
ab (mediated effect)	0.04	0.02	–	–0.00 to 0.09
ab/c (IL-6 percentage mediated) = no mediation				

Note: Significant results are reported in bold.

All models are adjusted for age, gender, body mass index and smoking.

Abbreviations: A $\beta_{1-40}$  and A $\beta_{1-42}$ : amyloid beta 1-40 and 1-42; A $\beta_{42:40}$ : amyloid beta 1-40 and 1-42 ratio; hs-CRP: high-sensitivity C-reactive protein; IL-6: interleukin 6.

willing to participate in the study compared to less sociable and introverted subjects (Wacholder, Silverman, McLaughlin, & Mandel, 1992). Therefore, potentially this type of controls could show better oral and general health as well as less systemic inflammation than the general population. Although a full cognitive assessment including specific cognitive tests was not done in these participants, none of them showed clinical cognitive dysfunction (confirmed by subjective cognitive examination recording slowness of thought, inappropriateness and mood) (Kipps & Hodges, 2005) or neuroimaging findings potentially associated with any evident form of dementia (e.g. brain damage, cortical/subcortical atrophy, hydrocephaly and cerebral small vessel disease). Future studies including cognitive healthy subjects confirmed with robust cognitive tests are needed to confirm our results. Another limitation to be considered was the use of traditional ELISA technique to measure blood A $\beta$  peptides. New ultrasensitive technologies (i.e. immunoaffinity-based assays) now available in the market such as immunomagnetic reduction (IMR) and single molecule array (SIMOA) must be considered in future studies to overcome the challenges of detection encountered using ELISA-based technique. Both IMR and SIMOA, although promising, still need validation in different populations by means of multicentre studies (Lue, Guerra, & Walker, 2017). Lastly, readers should interpret the present results with caution. AD pathogenesis is very complex. The amyloid hypothesis might not explain all cases of AD. In fact, cognitively normal individuals can have A $\beta$  deposits and also AD patients can present very few A $\beta$  deposits (Edison et al., 2007; Li et al., 2008). Distribution and extension of senile plaques are sometimes similar in patients with dementia compared to subjects with preserved cognitive function (Davis, Schmitt, Wekstein, & Markesbery, 1999; Fagan et al., 2009). Furthermore, A $\beta$  peptide accumulation alone represents an extremely early event in the amyloid cascade leading to neurodegeneration (Ricciarelli & Fedele, 2017). Further studies including AD patients

with and without periodontitis are needed to demonstrate whether periodontitis can feature as an early therapeutic target for AD.

In summary, it can be concluded that periodontitis is associated with high serum levels of A $\beta$  peptides. This finding could be due to enhanced systemic inflammation observed in patients with periodontitis. Further longitudinal evidence from cohort studies using different populations is warranted to confirm our preliminary results. Future research investigating the potential role of periodontitis in the neurodegenerative process is needed.

#### CONFLICT OF INTEREST

The authors have stated explicitly that there are no conflicts of interest in connection with this article.

#### ORCID

Yago Leira  <https://orcid.org/0000-0001-5027-7276>

Francesco D'Aiuto  <https://orcid.org/0000-0001-8654-935X>

Juan Blanco  <https://orcid.org/0000-0001-9251-513X>

#### REFERENCES

- Ainamo, J., & Bay, I. (1975). Problems and proposals for recording gingivitis and plaque. *International Dental Journal*, 25, 229–235.
- Amar, S., Gokce, N., Morgan, S., Loukideli, M., Van Dyke, T. E., & Vita, J. A. (2003). Periodontal disease is associated with brachial artery endothelial dysfunction and systemic inflammation. *Arteriosclerosis, Thrombosis and Vascular Biology*, 23, 1245–1249. <https://doi.org/10.1161/01.ATV.0000078603.90302.4A>
- Ameijera, P., Leira, Y., Domínguez, C., Leira, R., & Blanco, J. (2019). Association between periodontitis and chronic migraine: A case-control study. *Odontology*, 107, 90–95. <https://doi.org/10.1007/s10266-018-0360-7>
- Ardila, C. M., & Guzmán, I. C. (2015). Comparison of serum amyloid A protein and C-reactive protein levels as inflammatory markers in periodontitis. *Journal of Periodontal and Implant Science*, 45, 14–22. <https://doi.org/10.5051/jpis.2015.45.1.14>

- D'Aiuto, F., Parkar, M., Andreou, G., Suvan, J., Brett, P. M., Ready, D., & Tonetti, M. S. (2004). Periodontitis and systemic inflammation: Control of the local infection is associated with a reduction in serum inflammatory markers. *Journal of Dental Research*, *83*, 156–160. <https://doi.org/10.1177/154405910408300214>
- Davis, D. G., Schmitt, F. A., Wekstein, D. R., & Markesbery, W. R. (1999). Alzheimer neuropathologic alterations in aged cognitively normal subjects. *Journal of Neuropathology and Experimental Neurology*, *58*, 376–388. <https://doi.org/10.1097/00005072-199904000-00008>
- De Miguel, C., Rudemiller, N. P., Abais, J. M., & Mattson, D. L. (2015). Inflammation and hypertension: New understandings and potential therapeutic targets. *Current Hypertension Reports*, *17*, 507. <https://doi.org/10.1007/s11906-014-0507-z>
- Dominy, S. S., Lynch, C., Ermini, F., Benedyk, M., Marczyk, A., Konradi, A., ... Potempa, J. (2019). *Porphyromonas gingivalis* in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. *Science Advances*, *5*, eaau3333. <https://doi.org/10.1126/sciadv.aau3333>
- Edison, P., Archer, H. A., Hinz, R., Hammers, A., Pavese, N., Tai, Y. F., ... Brooks, D. J. (2007). Amyloid, hypometabolism, and cognition in Alzheimer disease: An [11C]PIB and [18F]FDG PET study. *Neurology*, *68*, 501–508. <https://doi.org/10.1212/01.wnl.0000244749.20056.d4>
- Eke, P. I., Page, R. C., Wei, L., Thornton-Evans, G., & Genco, R. J. (2012). Update of the case definitions for population-based surveillance of periodontitis. *Journal of Periodontology*, *83*, 1449–1454. <https://doi.org/10.1902/jop.2012.110664>
- Ellulu, M. S., Patimah, I., Khaza'ai, H., Rahmat, A., & Abed, Y. (2017). Obesity and inflammation: The linking mechanism and the complications. *Archives of Medical Science*, *13*, 851–863. <https://doi.org/10.5114/aoms.2016.58928>
- Fagan, A. M., Mintun, M. A., Shah, A. R., Aldea, P., Roe, C. M., Mach, R. H., ... Holtzman, D. M. (2009). Cerebrospinal fluid tau and ptau(181) increase with cortical amyloid deposition in cognitively normal individuals: Implications for future clinical trials of Alzheimer's disease. *EMBO Molecular Medicine*, *1*, 371–380. <https://doi.org/10.1002/emmm.200900048>
- Gil-Montoya, J. A., Barrios, R., Santana, S., Sanchez-Lara, I., Pardo, C. C., Fornieles-Rubio, F., ... Burgos, J. S. (2017). Association between periodontitis and amyloid  $\beta$  peptide in elderly people with and without cognitive impairment. *Journal of Periodontology*, *88*, 1051–1058. <https://doi.org/10.1902/jop.2017.170071>
- Hayes, A. F. (2013). *Introduction to mediation, moderation, and conditional process analysis: A regression-based approach*. New York, NY: The Guilford Press.
- Holmer, J., Eriksdotter, M., Schultzberg, M., Pussinen, P. J., & Buhlin, K. (2018). Association between periodontitis and risk of Alzheimer's disease, mild cognitive impairment and subjective cognitive decline: A case-control study. *Journal of Clinical Periodontology*, *45*, 1287–1298. <https://doi.org/10.1111/jcpe.13016>
- Hujoel, P. P., White, B. A., García, R. I., & Listgarten, M. A. (2001). The dentogingival epithelial surface area revisited. *Journal of Periodontal Research*, *36*, 48–55. <https://doi.org/10.1034/j.1600-0765.2001.00011.x>
- Iadecola, C., Park, L., & Capone, C. (2009). Threats to the mind: Aging, amyloid, and hypertension. *Stroke*, *40*, 40–44. <https://doi.org/10.1161/STROKEAHA.108.533638>
- Ide, M., Harris, M., Stevens, A., Sussams, R., Hopkins, V., Culliford, D., ... Holmes, C. (2016). Periodontitis and cognitive decline in Alzheimer's disease. *PLoS ONE*, *11*, e0151081. <https://doi.org/10.1371/journal.pone.0151081>
- Kamer, A. R., Pirraglia, E., Tsui, W., Rusinek, H., Vallabhajosula, S., Mosconi, L., ... de Leon, M. J. (2015). Periodontal disease associates with higher brain amyloid load in normal elderly. *Neurobiology of Aging*, *36*, 627–633. <https://doi.org/10.1016/j.neurobiolaging.2014.10.038>
- Kipps, C. M., & Hodges, J. R. (2005). Cognitive assessment for clinicians. *Journal of Neurology, Neurosurgery and Psychiatry*, *76*, 22–30. <https://doi.org/10.1136/jnnp.2004.059758>
- Kubota, T., Maruyama, S., Abe, D., Tomita, T., Morozumi, T., Nakasone, N., ... Yoshie, H. (2014). Amyloid beta (A $\beta$ ) precursor protein expression in human periodontitis-affected gingival tissues. *Archives of Oral Biology*, *59*, 586–594. <https://doi.org/10.1016/j.archoralbio.2014.03.004>
- Lee, Y. H., Tharp, W. G., Maple, R. L., Nair, S., Permana, P. A., & Pratley, R. E. (2008). Amyloid precursor protein expression is upregulated in adipocytes in obesity. *Obesity*, *16*, 1493–14500. <https://doi.org/10.1038/oby.2008.267>
- Leira, Y., & Blanco, J. (2018). Brain natriuretic peptide serum levels in periodontitis. *Journal of Periodontal Research*, *53*, 575–581. <https://doi.org/10.1111/jre.12547>
- Leira, Y., Domínguez, C., Seoane, J., Seoane-Romero, J., Pías-Peleteiro, J. M., Takkouche, B., ... Aldrey, J. M. (2017). Is periodontal disease associated with Alzheimer's disease? A systematic review with meta-analysis. *Neuroepidemiology*, *48*, 21–31. <https://doi.org/10.1159/000458411>
- Leira, Y., Iglesias-Rey, R., Gómez-Lado, N., Aguiar, P., Campos, F., D'Aiuto, F., ... Sobrino, T. (2019). *Porphyromonas gingivalis* lipopolysaccharide-induced periodontitis and serum amyloid-beta peptides. *Archives of Oral Biology*, *99*, 120–125. <https://doi.org/10.1016/j.archoralbio.2019.01.008>
- Leira, Y., Rodríguez-Yáñez, M., Arias, S., López-Dequidt, I., Campos, F., Sobrino, T., ... Blanco, J. (2019a). Periodontitis is associated with systemic inflammation and vascular endothelial dysfunction in patients with lacunar infarct. *Journal of Periodontology*, *90*, 465–474. <https://doi.org/10.1002/JPER.18-0560>
- Leira, Y., Rodríguez-Yáñez, M., Arias, S., López-Dequidt, I., Campos, F., Sobrino, T., ... Blanco, J. (2019b). Periodontitis as a risk indicator and predictor of poor outcome for lacunar infarct. *Journal of Clinical Periodontology*, *46*(1), 20–30. <https://doi.org/10.1111/jcpe.13032>
- Li, Y., Rinne, J. O., Mosconi, L., Pirraglia, E., Rusinek, H., Desanti, S., ... de Leon, M. J. (2008). Regional analysis of FDG and PIB-PET images in normal aging, mild cognitive impairment, and Alzheimer's disease. *European Journal of Nuclear Medicine and Molecular Image*, *35*, 2169–2181. <https://doi.org/10.1007/s00259-008-0833-y>
- Linden, G. J., McClean, K., Young, I., Evans, A., & Kee, F. (2008). Persistently raised C-reactive protein levels are associated with advanced periodontal disease. *Journal of Clinical Periodontology*, *35*, 741–747. <https://doi.org/10.1111/j.1600-051X.2008.01288.x>
- Loos, B. G., Craandijk, J., Hoek, F. J., Wertheim-van Dillen, P. M., & van der Velden, U. (2000). Elevation of systemic markers related to cardiovascular diseases in the peripheral blood of periodontitis patients. *Journal of Periodontology*, *71*, 1528–1534. <https://doi.org/10.1902/jop.2000.71.10.1528>
- Lue, L.-F., Guerra, A., & Walker, D. G. (2017). Amyloid beta and Tau as Alzheimer's disease blood biomarkers: Promise from new technologies. *Neurology and Therapy*, *6*, 25–36. <https://doi.org/10.1007/s40120-017-0074-8>
- Magalingam, K. B., Radhakrishnan, A., Ping, N. S., & Haleagrahara, N. (2018). Current concepts of neurodegenerative mechanisms in Alzheimer's disease. *BioMed Research International*, *3740461*, <https://doi.org/10.1155/2018/3740461>
- Marsland, A. L., McCaffery, J. M., Muldoon, M. F., & Manuck, S. B. (2010). Systemic inflammation and the metabolic syndrome among middle-aged community volunteers. *Metabolism*, *59*, 1801–1808. <https://doi.org/10.1016/j.metabol.2010.05.015>
- Murphy, M. P., & LeVine, H. 3rd (2010). Alzheimer's disease and the amyloid-beta peptide. *Journal of Alzheimer's Disease*, *19*, 311–323. <https://doi.org/10.3233/JAD-2010-1221>
- Nesse, W., Abbas, F., van der Ploeg, I., Spijkervet, F. K., Dijkstra, P. U., & Vissink, A. (2008). Periodontal inflamed surface area: Quantifying

- inflammatory burden. *Journal of Clinical Periodontology*, 35, 668–673. <https://doi.org/10.1111/j.1600-051X.2008.01249.x>
- Niwa, K., Carlson, G. A., & Iadecola, C. (2000). Exogenous A beta1-40 reproduces cerebrovascular alterations resulting from amyloid precursor protein overexpression in mice. *Journal of Cerebral Blood Flow and Metabolism*, 20, 1659–1668. <https://doi.org/10.1097/00004647-200012000-00005>
- Niwa, K., Younkin, L., Ebeling, C., Turner, S. K., Westaway, D., Younkin, S., ... Iadecola, C. (2000). Abeta 1-40-related reduction in functional hyperemia in mouse neocortex during somatosensory activation. *Proceedings of the National Academy of Sciences of the United States of America*, 97, 9735–9740. <https://doi.org/10.1073/pnas.97.17.9735>
- Noack, B., Genco, R. J., Trevisan, M., Grossi, S., Zambon, J. J., & De Nardin, E. (2001). Periodontal infections contribute to elevated systemic C-reactive protein level. *Journal of Periodontology*, 72, 1221–1227. <https://doi.org/10.1902/jop.2000.72.9.1221>
- Paraskevas, S., Huizinga, J. D., & Loos, B. G. (2008). A systematic review and meta-analyses on C-reactive protein in relation to periodontitis. *Journal of Clinical Periodontology*, 35, 277–290. <https://doi.org/10.1111/j.1600-051X.2007.01173.x>
- Perry, V. H. (2004). The influence of systemic inflammation on inflammation in the brain: Implications for chronic neurodegenerative disease. *Brain, Behaviour, and Immunity*, 18, 407–413. <https://doi.org/10.1016/j.bbi.2004.01.004>
- Pitiphat, W., Savetsilp, W., & Wara-Aswapati, N. (2008). C-reactive protein associated with periodontitis in a Thai population. *Journal of Clinical Periodontology*, 35, 120–125. <https://doi.org/10.1111/j.1600-051X.2007.01179.x>
- Razay, G., Vreugdenhil, A., & Wilcock, G. (2007). The metabolic syndrome and Alzheimer disease. *Archives of Neurology*, 64, 93–96. <https://doi.org/10.1001/archneur.64.1.93>
- Ricciarelli, R., & Fedele, E. (2017). The amyloid cascade hypothesis in Alzheimer's disease: It's time to change our mind. *Currents in Neuropharmacology*, 15, 926–935. <https://doi.org/10.2174/1570159X15666170116143743>
- Shah, N. S., Vidal, J. S., Masaki, K., Petrovitch, H., Ross, G. W., Tilley, C., ... Launer, L. J. (2012). Midlife blood pressure, plasma  $\beta$ -amyloid, and the risk for Alzheimer disease: The Honolulu Asia Aging Study. *Hypertension*, 59, 780–786. <https://doi.org/10.1161/HYPERTENSI.0NAHA.111.178962>
- Slade, G. D., Offenbacher, S., Beck, J. D., Heiss, G., & Pankow, J. S. (2000). Acute-phase inflammatory response to periodontal disease in the US population. *Journal of Dental Research*, 79, 49–57. <https://doi.org/10.1177/00220345000790010701>
- Sproston, N. R., & Ashworth, J. J. (2018). Role of C-reactive protein at sites of inflammation and infection. *Frontiers of Immunology*, 9, 754. <https://doi.org/10.3389/fimmu.2018.00754>
- Takeuchi, K., Ohara, T., Furuta, M., Takeshita, T., Shibata, Y., Hata, J., ... Ninomiya, T. (2017). Tooth loss and risk of dementia in the community: The Hisayama Study. *Journal of the American Geriatrics Society*, 65, 95–100. <https://doi.org/10.1111/jgs.14791>
- Tiwari, S., Atluri, V., Kaushik, A., & Yndart, A. (2019). Alzheimer's disease: Pathogenesis, diagnostics, and therapeutics. *International Journal of Nanomedicine*, 14, 5541–5554. <https://doi.org/10.2147/IJN.S200490>
- Von Elm, E., Altman, D. G., Egger, M., Pocock, S. J., Gøtzsche, P. C., Vandenbroucke, J. P., & STROBE Initiative (2008). The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: Guidelines for reporting observational studies. *Journal of Clinical Epidemiology*, 61, 344–349. <https://doi.org/10.1016/j.jclinepi.2007.11.008>
- Wacholder, S., Silverman, D. T., McLaughlin, J. L., & Mandel, J. S. (1992). Selection of controls in cases-control studies. II. Type of controls. *American Journal of Epidemiology*, 135, 1029–1041. <https://doi.org/10.1093/oxfordjournals.aje.a116397>
- Wang, R. P., Ho, Y. S., Leung, W. K., Goto, T., & Chang, R. C. (2019). Systemic inflammation linking chronic periodontitis and cognitive decline. *Brain, Behaviour, and Immunity*, 81, 63–73. <https://doi.org/10.1016/j.bbi.2019.07.002>
- Wu, Z., Ni, J., Liu, Y., Teeling, J. L., Takayama, F., Colcutt, A., ... Nakanishi, H. (2017). Cathepsin B plays a critical role in inducing Alzheimer's disease-like phenotypes following chronic systemic exposure to lipopolysaccharide from *Porphyromonas gingivalis* in mice. *Brain, Behaviour, and Immunity*, 65, 350–361. <https://doi.org/10.1016/j.bbi.2017.06.002>

## SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

**How to cite this article:** Leira Y, Carballo Á, Orlandi M, et al. Periodontitis and systemic markers of neurodegeneration: A case-control study. *J Clin Periodontol*. 2020;47:561–571. <https://doi.org/10.1111/jcpe.13267>